T-677 P.004/013 F-257

Appln SN: 09/921,992

Response to Office Action mailed 01/16/2004

Atty Docket: REN-00-084-US

Amendments to the Specification:

Please replace the paragraph starting on page 91 line 8 with the following amended paragraph:

Alignment of the *E. coli* and *Arabidopsis gcpE* proteins shows high similarity but also striking differences. The first 75 amino acid residues of the *Arabidopsis* sequence constitute a region that is not present in the bacterial counterpart. A transit peptide for plastids is predicted at this region with the ChloroP V1.0 program accessible at the Center for Biological Sequences, University of Denmark web site and described by Emanuelsson, et al. (*Protein Science* 8:978-984, (1999)) www.ebs.dtu.dk/servicee/ChloroP/ (Score 0.53295). According to this program, the processing site of the transit peptide would be located between Arg38 and Ser39 (CS-score 2.392). *In vivo* import experiments to chloroplasts demonstrated that the N-terminal region of the *Arabidopsis* protein is a functional transit peptide for plastids.

Please replace the paragraph starting on page 100 line 1 with the following amended paragraph:

Upon identification of the Escherichia coli gcpE gene as involved in the trunk line of the MEP pathway for isoprenoid biosynthesis, the available databases are searched for plant homologs. As described in Example 4, clone 135H1 (Genbank accession number T46582) is identified as containing an Arabidopsis thaliana cDNA encoding a protein with homology to the product of the bacterial gcpE gene. As shown in Figure 4, however, the putative Arabidopsis GCPE protein (SEQ ID NO: 79), contains several domains that are absent from the E. coli protein (SEQ ID NO: 78). Identical residues are in black boxes and conservative changes in grey boxes. Gaps are indicated with dots. The predicted cleavage site for the plastidial targeting peptide (according to the ChloroP program, accessible at the Center for Biological Sequences, University of Denmark web site and described by Emanuelsson, et al. (Protein Science 8:978-984, (1999)); genome obseduedk/services/chlorop) is indicated with an arrow (see Figure 4).

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Please replace the paragraph starting on page 108 line 9 with the following amended paragraph:

The multiple gene construct contains the gcpE gene and one or more genes for other MEP pathway proteins, including, but not limited to: a ygbB gene; a ygbP gene; a ychB gene; a yfgA gene; a yfgB gene; a bifunctional prephenate dehydrogenase such as the E. herbicola or E. coli tyrA gene (Xia et al., J. Gen. Microbiol., 138:1309-1316, 1992), a phytylprenyltransferase such as the slr1736 gene (in Cyanobase, a web accessible database maintained by Kazusa DNA Research Institute, Kisarazu, JAPAN www.kazusa.or.jp/cyanobase) or the ATPT2 gene (Smith et al., Plant J., 11:83-92, 1997), a deoxyxylulose synthase such as the E. coli dxs gene (Lois et al., PNAS, 95(5):2105-2110, 1998), a deoxyxylulose reductoisomerase such as the dxr gene (Takahashi et al., PNAS, 95(17):9879-9884, 1998), an Arabidopsis thaliana HPPD gene (Norris et al., Plant Physiol., 117:1317-1323, 1998), an Arabidopsis thaliana GGPPS gene (Bartley and Scolnik, Plant Physiol., 104:1469-1470, 1994), a transporter such as the AANT1 gene (Saint Guily, et al., Plant Physiol., 100(2):1069-1071, 1992), a GMT gene (WO 00/32757, WO 00/10380), an MT1 gene, a tocopherol cyclase such as the slr1737 gene (in Cyanobase) or its Arabidopsis ortholog, an isopentenyl diphosphate isomerase (IDI) gene, and an antisense construct for homogentisic acid dioxygenase (Sato et al., J. DNA Res., 7(1):31-63, 2000).